



TRAIL-enhancing activity of Erythrinan alkaloids from *Erythrina velutina*

Masaaki Ozawa^a, Tadahiro Etoh^b, Masahiko Hayashi^b, Kanki Komiyama^c, Akio Kishida^a, Ayumi Ohsaki^{a,*}

^a Institute of Biomaterials and Bioengineering, Tokyo Medical and Dental University, 2-3-10, Surugadai, Kanda, Chiyoda-ku, Tokyo 101-0062, Japan

^b Iwaki Meisei University, 5-5-1, Iino, Chuo-dai, Iwaki, Fukushima 970-8551, Japan

^c The Kitasato Institute, 5-9-1, Shirokane, Minato-ku, Tokyo 108-8642, Japan

ARTICLE INFO

Article history:

Received 6 August 2008

Revised 6 October 2008

Accepted 25 October 2008

Available online 31 October 2008

Keywords:

TRAIL

Enhanced activity

Erythrinan alkaloid

Medicinal plant

Erythrina velutina

ABSTRACT

A new Erythrinan alkaloid (**1**), erythrodine N-oxide, was isolated from the seeds of *Erythrina velutina* together with seven known Erythrinan alkaloids (**2–7**, **9**) and an indole alkaloid (**8**). The structure of new compound (**1**) was elucidated by spectroscopic methods. Six of the nine compounds showed enhanced activity when combined with tumor necrosis factor-related apoptosis-inducing ligand (TRAIL). Erythrodine (**4**) did not show cytotoxic activity by itself, but exhibited significant cytotoxicity when combined with TRAIL.

© 2008 Elsevier Ltd. All rights reserved.

In search for biological active compounds derived from tropical medicinal plants,¹ we investigated the constituents of the seeds of *Erythrina velutina* Willd (Leguminosae).² This plant is commonly called 'Mulungu' in Brazil, and its bark is used for sedation, hypnogenesis, the control of convulsions, and nervous coughs in the northern region of Brazil.³ In a previous paper, we described the isolation and characterization of an indole derivative, hypaphorine (**8**), and its sleep-inducing effect in normal mice.⁴ In this paper, we describe the isolation and structural characterization of a new Erythrinan alkaloid, erythrodine N-oxide (**1**), together with eight known alkaloids (**2–9**) from the seeds of *E. velutina*, and these isolated compounds were evaluated regarding their TRAIL enhanced activity (Fig. 1).⁵

Tumor necrosis factor (TNF)-related apoptosis-inducing ligand (TRAIL) is an important member of the TNF superfamily.⁶ TRAIL selectively induces the apoptosis of a wide variety of cancer and transformed cells without damaging most normal cells. However, the clinical application of TRAIL in cancer therapy is limited as many cancer cells have been identified as resistant to the cytotoxicity of TRAIL.

Therefore, natural products, which have synergistic activity with TRAIL but minimal toxicity would be promising tools for cancer therapy. Recent studies on natural products showing enhanced activity identified flavonoidal compounds.⁷ However, the present report focuses on alkaloidal compounds from natural re-

sources, possibility leading to the expansion of promising candidates.

The seeds of *E. velutina* (954 g) were crushed and then extracted with MeOH. The MeOH-soluble materials (19.0 g) were succes-

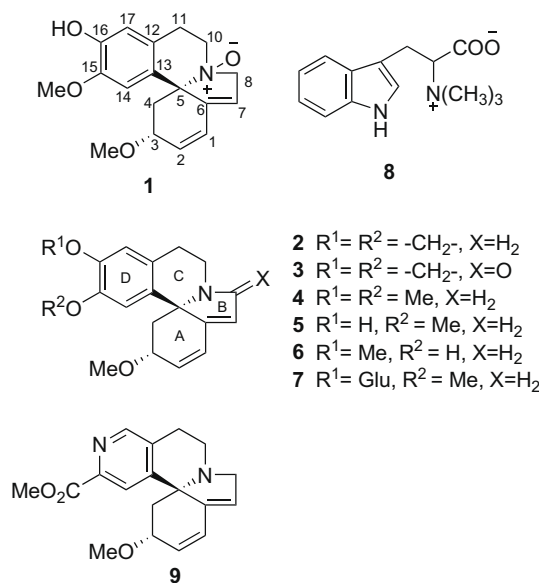


Figure 1. Chemical structures of compounds **1–9**.

* Corresponding author. Tel./fax: +81 3 5280 8153.

E-mail address: a-ohsaki.fmd@tmd.ac.jp (A. Ohsaki).

sively partitioned between petroleum ether, EtOAc, and 3% aqueous tartaric acid. Water-soluble materials were adjusted to pH 10 with Na_2CO_3 and then partitioned between CHCl_3 (6.07 g) and n -BuOH (3.48 g), to obtain the alkaloidal portions.

The CHCl_3 -soluble materials (3.11 g) were separated in a silica gel column (CHCl_3 –MeOH, 100:0 \rightarrow 0:100, Wako-gel C-300, Wako Ltd) to obtain 20 fractions (F1–F20). The second fraction (F2, 131 mg) was further separated using a silica gel column (EtOAc–MeOH, 99:1) to isolate erythraline (**2**, 9.0 mg),⁸ 8-oxo-erythraline (**3**, 87.0 mg),⁹ and erysotrine (**4**, 5.0 mg),⁸ F4 (11 mg) was separated using silica gel sheets (EtOAc–MeOH, 85:15, Si60-F254S, aluminum sheets, Merck) to give erysodine (**5**, 4.5 mg)¹⁰ and erysovine (**6**, 3.3 mg),⁸ and the last fraction (F-20, 349 mg) was separated using a silica gel column (MeOH/acetone, 99:1) to afford glycoerysodine (**7**, 5.0 mg)¹⁰ and hypaphorine (**8**, 44.0 mg).¹¹

On the other hand, the second extracts of constituents were extracted by 80% aqueous MeOH to obtain soluble materials (24.0 g), and these were partitioned using the same method to obtain alkaloidal fractions of CHCl_3 -soluble (C-2, 440 mg,) and n -BuOH-soluble (B-2, 4.86 g) materials. The C-2 fraction was separated using an amino silica gel column (Hexane–EtOAc, 90:10 \rightarrow 0:100, Chromatorex-NH, Fuji Silysia Ltd) to give 11 fractions (F₂-1–11). The third fraction (F₂-3, 26.4 mg) was purified by employing a silica gel column to obtain erymelanthine (**9**, 1.4 mg).¹² Furthermore, the B-2 extracts were separated using an amino silica gel column (CHCl_3 –MeOH, 100:0 \rightarrow 90:10), followed by a silica gel column (CHCl_3 –MeOH, 100:0 \rightarrow 90:10) to give a new Erythrinan compound (**1**, 2.6 mg), erysodine N-oxide.

Compound **1** [$[\alpha]_D^{25} +172.0$ (c 0.061, CHCl_3)] was obtained as a colorless, amorphous solid. Its molecular formula was determined as $\text{C}_{18}\text{H}_{21}\text{NO}_4$ by HREIMS [m/z 315.1456 (M)⁺, calcd for 315.1471] and the IR spectrum implied the presence of a hydroxyl group (3396 cm^{-1}), an aromatic moiety and a conjugate olefin (1647 , 1609 , 1516 , and 1457 cm^{-1}), respectively. The UV spectrum showed λ_{max} (MeOH) at 204 ($\log \epsilon$ 5.69), 221 ($\log \epsilon$ 5.18), and 234 ($\log \epsilon$ 5.24). The ^1H and ^{13}C NMR¹³ and HMQC data for **1** indicated the presence of four methylenes, one sp^3 oxy-methine, one sp^3 quaternary carbon, three sp^2 methines, one sp^2 quaternary carbon, two aromatic quaternary carbons, two aromatic methine carbons, two aromatic oxy-quaternary carbons, and two methoxy groups. With two double bonds and an aromatic ring being accounted for six of nine unsaturations, it was concluded that **1** contains four rings including one aromatic ring. The ^1H – ^1H COSY spectrum revealed connectivities of C-1 to C-4, C-7 to C-8, and C-10 to C-11 (Fig. 2). HMBC correlations (Fig. 2) were observed: H-17/C-11, 12 and 16, H-14/C-5 (δ_{C} 82.94; a spiro-carbon), 13, 15, H-11/C-10, 12, 13, 17, H-10/C-5, 11, H-8/C-6, 7, H-7/C-5, 6, H-1/C-2, 6, H-2/C-1, 3, H-3/C-4,

5 and H-4/C-3, 5, 6. These correlations showed that **1** was a typical Erythrinan alkaloid having a characteristic spiro-carbon positioned in center of rings of A, B, and C with two olefins of $\Delta^{1,2}$ (δ_{H} 6.60 dd, δ_{C} 125.2/ δ_{H} 6.18 br d, δ_{C} 134.2) and $\Delta^{6,7}$ (δ_{C} 139.1/ δ_{H} 5.68 br s, δ_{C} 117.2) and one aromatic hydroxyl group attached to the C-16 position. Further, two methoxy groups were attached to C-3 and C-15, as deduced from the HMBC correlations of H₃–3-OMe and H₃–15-OMe, to each methoxy-bearing carbons, respectively. Finally, the molecular formula of **1** indicated the presence of one more oxygen atom. On ^{13}C NMR, the spiro-carbon (C-5) and methylene carbons C-8 and C-10 were markedly deshielded by 16.1, 18.7, and 15.7 ppm in comparison with those of erysodine (**3**), respectively. These facts suggested that the nitrogen atom was oxygenated. The direction of the oxygen atom of N-oxide might be α -facing due to the downfield shifts of H-4 α (δ_{H} 3.24). The NOESY correlations (Fig. 3) of H₃–15-OMe/H-14, H-14/H-4 β , H-14/H-3, and H₃–15-OMe/H-4 β suggested that the methoxy group was placed at C-15. Another methoxy group located at C-3 had an α -configuration from the NOESY correlations of H₃–3-OMe/H-2, H₃–3-OMe/H-3, and H₃–3-OMe/H-4 α . Other NOESY correlations were satisfied with the relative stereochemistry of the new compound **1** depicted in Figure 3. Therefore, compound **1** was assigned as erysodine N-oxide (**1**) (Fig. 2). The CD spectrum of **1** showed a Cotton effect at λ_{max} (MeOH) 238 nm ($\Delta\epsilon$ +25.1), which suggested that the absolute

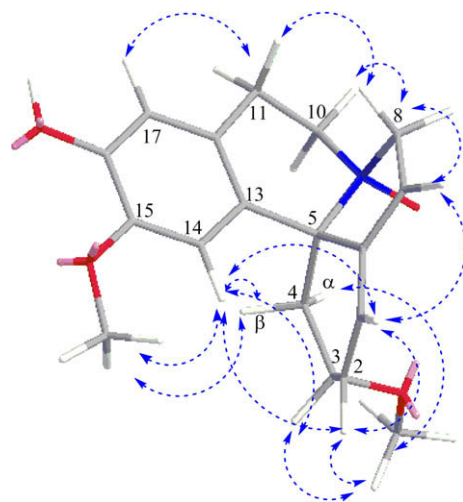


Figure 3. Selected NOESY correlations of compound **1**.

Table 1

IC_{50} ($\mu\text{g/mL}$) values of compounds (**1**–**9**) isolated from *E. velutina* with TRAIL for Jurcat cells.^a

Compound	No TRAIL ^b	With TRAIL ^c
1	>50 (2.4)	>50 (18.9)
2	23.0	5.0
3	>50 (37)	22.0
4	>50 (45)	7.2
5	10.8	4.9
6	24.0	14.2
7	>50 (4.7)	44.0
8	>50 (5.8)	>50 (20.0)
9	>50 (1.1)	>50 (24.2)
Apigenin ^d	30.0	6.7
Vincristine ^e	3.1 (ng/mL)	ND

^a 0.125 $\mu\text{g/mL}$ of TRAIL caused a 21.0% growth inhibition of Jurcat cells.

^b Numbers in parenthesis indicate the growth inhibitory % at a dose of 50 $\mu\text{g/mL}$.

^c The results are expressed relative to the cell proliferation rate at 0.125 $\mu\text{g/mL}$.

^d Positive control as TRAIL enhancement.

^e Positive control for Jurcat cells.

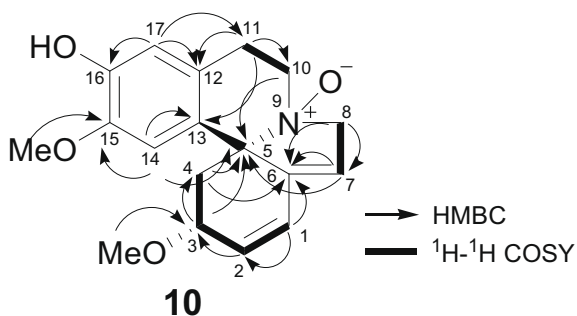


Figure 2. ^1H – ^1H COSY and selected HMBC correlations of compound **1**.

configuration of the 3-methoxy group of **1** was *R*.¹⁴ Therefore, the absolute configuration of **1** was assigned as depicted.

We examined the enhancing effects of the various compounds isolated from the seeds of *E. velutina* on the cytotoxicity of TRAIL against Jurkat cells. The results of proliferation tests are shown in Table 1. Most of the compounds tested showed an enhanced cytotoxicity against Jurkat cells when combined with TRAIL. Erythraline (**2**), erysodine (**5**), and erysovine (**6**) showed IC₅₀ values of 23.0, 10.8, and 24.0 µg/mL, respectively, and their cytotoxicities were further enhanced by the addition of TRAIL, up to an IC₅₀ values of 5.0, 4.9, and 14.2 µg/mL, respectively. However, these compounds showed some cytotoxicity even if their activities were moderate in the absence of TRAIL. The other compounds tested, 8-oxo-erythraline (**3**), erysotrine (**4**), and grycoerysodine (**7**), showed promising data, namely, these compounds exhibit no cytotoxicity by themselves, but they act synergistically when combined with TRAIL. Erysotrine (**4**) showed the superior data when combined with TRAIL (0.125 µg/mL) among the samples tested, and exhibited no cytotoxicity (>50 µg/mL) by itself.

The results of Erythrinan alkaloids regarding the synergistically enhancing effect of TRAIL indicated the expansion of candidates beyond flavonoidal or related compounds.

Acknowledgment

We thank Dr. G. Hashimoto (Centro de Pesquisas de História Natural, São Paulo, Brazil) for the identification of *Erythrina velutina*.

References and notes

- Ohsaki, A.; Takashima, J.; Chiba, N.; Kawamura, M. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 1109.
- Hashimoto, G. *Illustrated Cyclopedia of Brazilian Medicinal Plants*; ABOC-SHA: Kamakura, 1996. p 679.
- Dantas, M. C.; De Oliveira, F. S.; Bandeira, S. M.; Batista, J. S.; Silva, C. D.; Alves, P. B.; Antonioli, A. R.; Marchioro, M. *J. Ethnopharmacol.* **2004**, *94*, 129.
- Ozawa, M.; Honda, K.; Nakai, I.; Kishida, A.; Ohsaki, A. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 3992.
- Takashima, J.; Ikeda, Y.; Komiyama, K.; Hayashi, M.; Kishida, A.; Ohsaki, A. *Chem. Pharm. Bull.* **2007**, *55*, 343.
- Wang, S.; El-Deiry, W. S. *Oncogene* **2003**, *22*, 8628.
- (a) Horinaka, M.; Yoshida, T.; Shiraishi, T.; Nakata, S.; Wakada, M.; Nakanishi, R.; Nishino, H.; Sakai, T. *Biochem. Biophys. Res. Commun.* **2005**, *333*, 833; (b) Horinaka, M.; Yoshida, T.; Shiraishi, T.; Nakata, S.; Wakada, M.; Sakai, T. *Mol. Cancer Ther.* **2006**, *5*, 945. and references therein.
- Chawla, A. S.; Chunchatprasert, S.; Jacson, A. H. *Org. Magn. Reson.* **1983**, *21*, 39.
- Mantle, P. G.; Laws, I.; Widdowson, D. A. *Phytochemistry* **1984**, *23*, 1336.
- Amer, M. E.; El-Masry, S. *J. Nat. Prod.* **1991**, *54*, 161.
- Kondo, K.; Nishi, J.; Kobayashi, J. *J. Nat. Prod.* **1994**, *54*, 161.
- Dagne, E.; Steglich, W. *Tetrahedron Lett.* **1983**, *24*, 5067.
- NMR data for **1**: ¹H NMR (CDCl₃, 500 MHz) δ 6.6 0 dd, *J* = 10.1, 2.1, H-1), 6.18 (br d, *J* = 10.1, H-2), 4.21 (br dd, *J* = 6.0, 11.6, H-3), 2.09 (dd, *J* = 6.0, 11.6, H-4), 3.24 (t, *J* = 11.6, H-4), 5.68 (br s, H-7), 4.13 (dd, *J* = 14.4, 3.3, H-8), 4.43 (br d, *J* = 14.4, H-8), 3.81–3.85 (m, H₂-10), 3.03–3.16 (m, H₂-11), 6.79 (s, H-14), 6.56 (s, H-17), 3.35 (s, H₃-3-OMe), 3.86 (s, H₃-15-OMe); ¹³C NMR (CDCl₃, 125 Hz), δ 125.2 (C-1), 134.2 (C-2), 75.7 (C-3), 31.2 (C-4), 82.9 (C-5), 139.1 (C-6), 117.2 (C-7), 70.9 (C-8), 59.7 (C-10), 27.2 (C-11), 120.0 (C-12), 129.0 (C-13), 112.6 (C-14), 147.6 (C-15), 146.3 (C-16), 110.3 (C-17), 56.0 (C-3-OMe), 55.9 (C-15-OMe).
- Tsuda, Y.; Sano, T. *The Alkaloids*; Academic Press Inc., 1996. Vol. 48, p 249.